

SYNTHESIS OF A BIOSYNTHETIC  
PRECURSOR OF OLEANDOMYCIN,  
8,8a-DEOXYOLEANDOLIDE  
(8-METHYLOLEANDOLIDE),  
FROM OLEANDOMYCIN†

Sir:

A 14-membered macrolide aglycone, 8,8a-deoxyoleandolide<sup>1)</sup> (8-methyloleandolide<sup>2)</sup> (**1**) has been known as a precursor in the biosynthesis of oleandomycin (**2**), which is a clinically important macrolide antibiotic. The isolation and determination of structure of this interesting substance **1** by Abbott group<sup>1)</sup> are among the most notable of developments in the field of the macrolide biosynthesis. Because of the biosynthetic significance and the extreme scarcity of the macrolide **1**, there has been considerable interest in the practical synthesis.

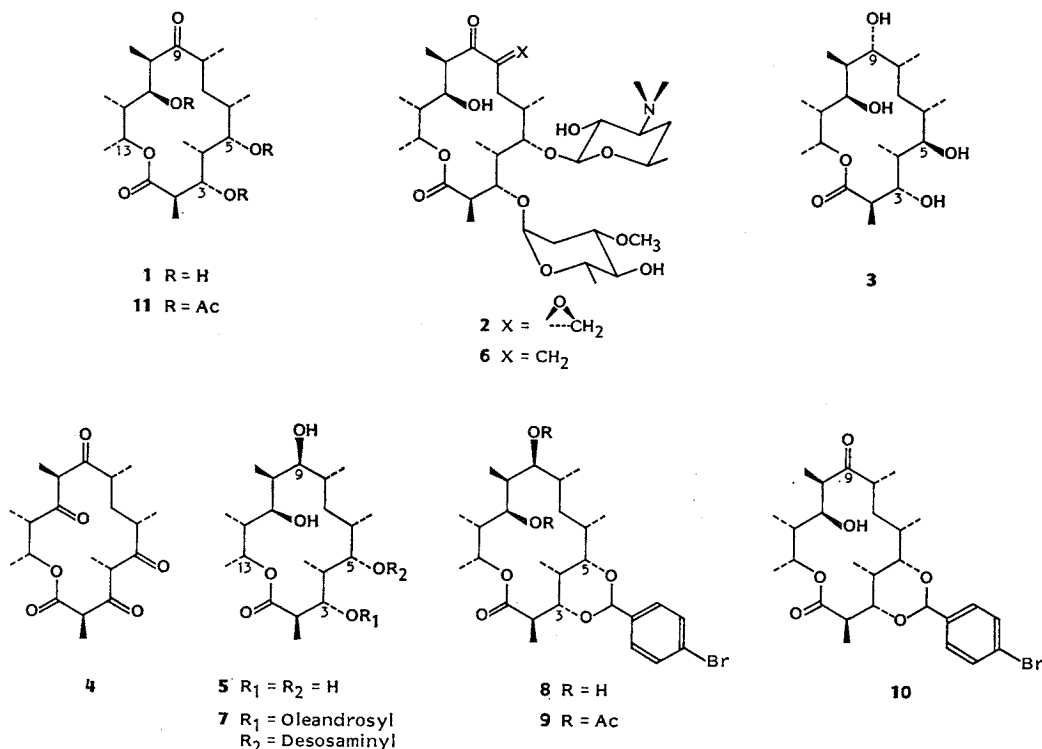
Very recently, we reported a novel enantioselective synthesis of the analogous aglycone, (5*R*,8*R*,9*R*)-9-dihydro-8-methyl-*epi*-oleandolide (**3**), which was incompatible with **1** only in the stereochemistry at the C-5 position, from the polyketide lactone **4** in order to chemically

simulate a probable biosynthetic pathway of the macrolide<sup>3)</sup>. The key intermediate **4** was effectively derived from a new aglycone of oleandomycin, (8*R*,9*S*)-9-dihydro-8-methyloleandolide (**5**)<sup>3)</sup>.

Herein, we describe an efficient synthesis of 8-methyloleandolide (**1**) by using the aglycone **5**.

The intermediary aglycone **5** [mp 99°C;  $[\alpha]_D^{25} +22^\circ$  (*c* 0.5, CHCl<sub>3</sub>)] was prepared in six steps by our method<sup>3)</sup> beginning with the formation of the exocyclic methylene **6** from oleandomycin **2** as follows: i) Treatment of **2** with CrCl<sub>2</sub> to give **6**; ii) successive stereospecific reductions with hydrogen and Raney Ni to give the (8*R*)-8-methyl compound, followed by NaBH<sub>4</sub> to give (8*R*,9*S*)-9-dihydro-8-methyloleandomycin (**7**); iii) removal of the sugar moieties by acid hydrolysis to give the deoleandrosyl compound, followed by reaction of the corresponding *N*-oxide with trimethylsilyl iodide. The overall yield from **2** was 56%.

Regioselective benzylidenation of **5** with *p*-bromobenzaldehyde dimethyl acetal (bp<sub>4</sub> 84°C) and a catalytic amount of *p*-toluenesulfonic acid in CH<sub>2</sub>Cl<sub>2</sub> at 5°C for 5 hours afforded the corresponding 3,5-*O*-protected product **8** in 92%



† Dedicated to the memory of Professor HAMA O UMEZAWA.

Table 1. Selected  $^1\text{H}$  NMR parameters (500 MHz,  $\text{CDCl}_3$ ) of **1** and **8**~**11**.

	Chemical shifts ( $\delta$ )				
	<b>1</b>	<b>8</b>	<b>9</b>	<b>10</b>	<b>11</b>
H-2	2.74	2.86	2.82	2.90	2.78
H-3	3.89	3.74	3.85	3.81	5.22
H-4	1.86	1.91	~2.38	2.11	2.29
H-5	3.99	3.86	3.99	3.93	4.79
H-6	2.01	2.37	~2.38	2.36	~2.1
H-8	2.62	1.52	~2.2	2.67	~2.75
H-9	—	3.12	4.76	—	—
H-10	~2.77	2.00	~2.2	~2.85	3.08
H-11	3.66	3.68	5.25	3.98	4.94
H-12	~1.72	1.65	1.68	1.66	1.81
H-13	5.49	5.58	4.93	5.64	5.19

	Coupling constants (Hz)				
	<b>1</b>	<b>8</b>	<b>9</b>	<b>10</b>	<b>11</b>
$J_{2,3}$	10.5	10.5	10.5	10.5	10.5
$J_{3,4}$	<1	<1	1.0	<1	1.5
$J_{4,5}$	2.5	1.0	1.0	1.0	6.0
$J_{5,6}$	4.5	6.0	6.0	6.0	2.0
$J_{6,9}$	—	9.0	11.0	—	—
$J_{9,10}$	—	3.0	3.0	—	—
$J_{10,11}$	2.0	2.0	0.5	2.5	2.0
$J_{11,12}$	10.5	10.5	10.5	10.5	10.5
$J_{12,13}$	1.5	1.0	1.0	1.5	1.5

yield, after silica gel column chromatography with hexane - EtOAc (2:1). Recrystallization from acetone - hexane gave cubics of **8**: MP 240°C;  $[\alpha]_D^{25} +15^\circ$  (*c* 0.58,  $\text{CHCl}_3$ ); field desorption mass spectrum (FD-MS)  $m/z$  (M+H) 542 and 544;  $^1\text{H}$  NMR (see Table 1). The structure was confirmed by the  $^1\text{H}$  NMR spectrum (Table 1) of the corresponding diacetate **9**: MP 200°C (needles after recrystallization from ether - hexane);  $[\alpha]_D^{30} -1.5^\circ$  (*c* 0.5,  $\text{CHCl}_3$ ); FD-MS  $m/z$  (M+H) 625 and 627.

Selective oxidation of **8** with pyridinium dichromate in  $\text{CH}_2\text{Cl}_2$  at 25°C for 3 hours gave exclusively the C-9 ketone **10** in 82% yield as a syrup, which was kept in hexane at 0°C to afford the amorphous solid: MP 117~125°C;  $[\alpha]_D^{30} -48^\circ$  (*c* 0.5,  $\text{CHCl}_3$ ); Rf 0.45 on TLC (hexane - EtOAc, 2:1); FD-MS  $m/z$  (M+H) 540 and 542;  $^1\text{H}$  NMR (see Table 1).

Removal of the benzylidene group without acetal formation between the C-5 hydroxyl and the C-9 carbonyl groups was best realized by hydrogenolysis<sup>4)</sup> with 3-atm hydrogen and Pd-black in EtOH to give 8-methyloleandolide (**1**)

as a syrup in 86% yield, after silica gel column chromatography with hexane - EtOAc (1:1). The syrup changed to needles of **1** by gradual evaporation of the  $\text{CHCl}_3$  solution around 0°C: MP 68~70°C;  $[\alpha]_D^{25} -47^\circ$  (*c* 0.5,  $\text{CHCl}_3$ ); Rf 0.43 on TLC (hexane - EtOAc 1:1); FD-MS  $m/z$  (M+H) 373; IR  $\nu_{\text{max}}^{\text{CHCl}_3} \text{ cm}^{-1}$  2980, 2940, 2880, 1700, 1457, 1374 and 1332;  $^1\text{H}$  NMR (see Table 1).

The compound **1** was further characterized by acetylation with  $\text{Ac}_2\text{O}$  and pyridine at 25°C for 24 hours to give, after silica gel column chromatography (benzene - EtOAc, 3:1), the crystalline triacetate **11** in 87% yield: MP 152~169°C (plates after recrystallization from  $\text{MeOH} - \text{H}_2\text{O}$ )<sup>1)</sup>; mp 174~176°C (needles after recrystallization from ether - hexane);  $[\alpha]_D^{28} -11^\circ$  (*c* 0.5,  $\text{CHCl}_3$ ); Rf 0.73 on TLC (benzene - EtOAc, 1:1); FD-MS  $m/z$  (M+H) 499; IR  $\nu_{\text{max}}^{\text{KBr}} \text{ cm}^{-1}$  1733 and 1700;  $^1\text{H}$  NMR (see Table 1).

These physico-chemical data of **1** and **11** were identical with those of naturally derived macrolides<sup>1)</sup>. Thus, pure synthetic macrolide **1** is now available in multigram amounts. Further

the detailed structure has been established unambiguously.

#### Acknowledgments

We are grateful to the Institute of Microbial Chemistry for the generous support of our program, and also thank Pfizer Taito Co., Ltd. for a generous gift of oleandomycin. Financial support by the Ministry of Education, Science and Culture (Grant-in-Aid Scientific Research) is gratefully acknowledged.

KUNIAKI TATSUTA  
YOSHIYUKI KOBAYASHI  
MITSUHIRO KINOSHITA

Department of Applied Chemistry,  
Keio University,  
3-14-1 Hiyoshi, Kohoku-ku,  
Yokohama 223, Japan

(Received January 13, 1987)

#### References

- 1) MARTIN, J. R.; R. S. EGAN, A. W. GOLDSTEIN, S. L. MUELLER, E. A. HIRNER & R. S. STANASZEK: 8,8a-Deoxyoleandolide: Elaborated by a blocked mutant of the erythromycin-producing organism *Streptomyces erythreus*. *J. Antibiotics* 27: 570~572, 1974
- 2) SAKAKIBARA, H. & S. ŌMURA: Chemical modification and structure-activity relationship of macrolides. *In* *Macrolide Antibiotics. Chemistry, Biology, and Practice. Ed., S. ŌMURA*, pp. 95~97, Academic Press, Inc., Tokyo, 1984
- 3) TATSUTA, K.; Y. KOBAYASHI, K. AKIMOTO & M. KINOSHITA: An enantioselective synthesis of a macrolide from the polyketide lactone derived from oleandomycin. *Chem. Lett.* 1987: 187~190, 1987
- 4) KINOSHITA, M.; M. ARAI, N. OHSAWA & M. NAKATA: Synthetic studies of erythronolide A through (9*S*)-9-dihydroerythronolide A. *Tetrahedron Lett.* 1986: 1815~1818, 1986